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Antiarrhythmic mechanisms of beta blocker therapy

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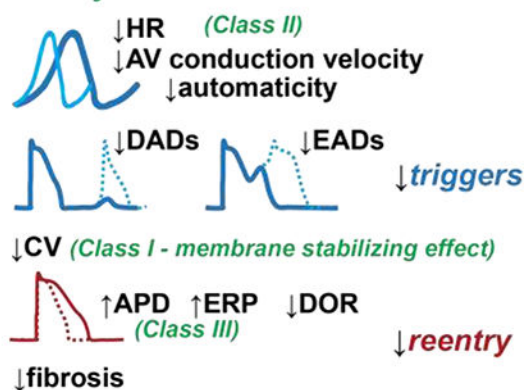
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Abstract

Sympathetic activity plays an important role in modulation of cardiac rhythm. Indeed, while exerting positive tropic effects in response to physiologic and pathologic stressors, β -adrenergic stimulation influences cardiac electrophysiology and can lead to disturbances of the heart rhythm and potentially lethal arrhythmias, particularly in pathological settings. For this reason, β -blockers are widely utilized clinically as antiarrhythmics. In this review, the molecular mechanisms of β -adrenergic action in the heart, the cellular and tissue level cardiac responses to β -adrenergic stimulation, and the clinical use of β -blockers as antiarrhythmic agents are reviewed. We emphasize the complex interaction between cardiomyocyte signaling, contraction, and electrophysiology occurring over multiple time- and spatial-scales during pathophysiological responses to β -adrenergic stimulation. An integrated understanding of this complex system is essential for optimizing therapies aimed at preventing arrhythmias.

Graphical Abstract

Antiarrhythmic effects of beta-blockers



Keywords

Autonomic drugs; arrhythmia; sympathetic stimulation; heart failure; beta-blockers

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1. Overview: the role of β -adrenergic activation in the heart

The sympathetic nervous system plays a key role in the neurohormonal control of cardiovascular function. It mediates neuronal and hormonal responses to fear, stress, or exercise by regulating cardiovascular function to meet the increasing demands of the body with a commensurate rapid increase in cardiac output (fight or flight response) (Cannon, 1915). The fight or flight response is initiated by release of norepinephrine (NE), from the cardiac sympathetic nerves, and epinephrine (Epi), from the adrenal medulla, which bind to β -adrenergic receptors (β -ARs) on cardiomyocytes. This triggers a signaling cascade leading to increase in cAMP and consequent PKA activation and phosphorylation of a myriad of targets, which orchestrate the physiological response of the heart: increase in heart rate and conduction velocity, and increased force of contraction and speed of relaxation (Bers, 2002). Although these responses are necessary to meet physical demands, excessive β -AR stimulation is also associated with electrophysiological abnormalities, leading to sometimes lethal disturbances of the cardiac rhythm, particularly in the setting of underlying cardiovascular disease (Ripplinger et al., 2016). Indeed, pharmacological agents with β -AR antagonism (class II antiarrhythmics or drugs with some β -AR blocking action, like amiodarone) are effective antiarrhythmics in many conditions (January et al., 2014; Al-Khatib et al., 2018). Here we review the intracellular pathways of β -adrenergic action, the mechanisms of cellular and tissue level regulation of cardiac electrical activity and arrhythmia, and the evidence for clinical antiarrhythmic indications for β -blocker therapy in cardiac diseases and arrhythmias, such as atrial fibrillation (AF), myocardial infarction (MI), and heart failure (HF). We suggest key questions and aspects that future experimentation and clinical studies should address to improve personalization of β -blockade therapy to varying arrhythmia types and patient groups.

2. Molecular mechanisms of β -adrenergic action

β -ARs are members of the G protein-coupled receptor superfamily. In the heart, at least two types of β -ARs are expressed, whereby β_1 -ARs account for the majority (~80%) and β_2 -ARs comprise ~20% of cardiac β -ARs (Bristow et al., 1986; Xiao, 2001). β_1 -ARs have been shown to initiate a cell-wide response, whereas β_2 -ARs are localized in caveolae and associated preferentially with L-type Ca^{2+} (Ca) channels (Xiao, 2001).

Binding of NE or Epi to β_1 -ARs activate stimulatory G proteins (G_s). The G_α subunit of the G_s protein ($G_{s\alpha}$) binds to and activates adenylyl cyclase (AC), which catalyzes the conversion of adenosine triphosphate (ATP) into the second messenger cyclic adenosine monophosphate (cAMP). cAMP also activates protein kinase A (PKA), which then phosphorylates several downstream targets in both contractile cells and in the conduction system, including L-type Ca channels, phospholamban (PLB), ryanodine receptors (RyRs), and myofilament proteins including troponin I (Figure 1). These proteins enable the coupling of cell excitation to contraction by increasing the amount of intracellular Ca at each systole (to augment contraction) and by decreasing the myofilament Ca sensitivity (to speed relaxation) (Bers, 2002). cAMP also binds directly to hyperpolarization-activated cyclic nucleotide gated (HCN) channels, predominantly expressed in cardiac nodal cells, to increase the pacemaker current I_f , which contributes to increased heart rate.

The documented effects of cardiac β_2 -AR activation are species-dependent, and vary with the developmental or pathophysiological state of the heart (reviewed in (Xiao, 2001)). β_2 -ARs couple with G_s at baseline, but in some conditions, they couple with the G inhibitory (G_i) proteins, whereby the latter releases the activated $G_i\alpha$ subunit to inhibit AC activity. The localization of β_2 -ARs in caveolae, closely associated with Ca channels (Figure 1), and the additional G_i pathway are thought to play a role in reshaping the spatiotemporal pattern of the G_s -AC-cAMP signaling and might have consequences on the kinetic (mismatch) between Ca and K^+ (K) channel responses to adrenergic activation. The G_i pathway also delivers G_s -independent signals, i.e., cell survival signals through a G_i - $G\beta\gamma$ -PI3K-Akt pathway that could be important in counteracting the detrimental effects of chronic β -AR activation (see also Section 5).

β_3 -ARs are expressed to much lesser degree, and their function in the heart has been poorly investigated (Cannavo & Koch, 2017). Whereas species differences have been reported, in human ventricle, β_3 -ARs mainly couple with $G_{s\alpha_i}$ proteins, and thus can counteract the effects of β_1 -AR and β_2 -AR activation (see also Section 5).

3. Cellular and tissue level β -adrenergic responses

3.1 Physiological responses

Sympathetic activation leads to increased heart rate (chronotropy), force of contraction (inotropy), speed of relaxation (lusitropy), and conduction (dromotropy). Positive chronotropy is mediated by phosphorylation-dependent changes in intracellular Ca handling as well as by cAMP-mediated increases in I_f (DiFrancesco & Tortora, 1991), which together accelerate diastolic depolarization in the sinoatrial (SA) node via the membrane and Ca coupled clocks (Lakatta & DiFrancesco, 2009), leading to faster impulse generation. Faster heart rates and K channel phosphorylation typically abbreviate cardiac repolarization (Bartos et al., 2015; Grandi et al., 2017), necessary to accommodate shorter cycle lengths, by counter-balancing the I_{CaL} increase necessary for enhancing contractility.

Increased myocardial contractility is mediated by cytosolic Ca increase, which increases the fraction of bound myosin and actin filaments, and is primarily due to enhancement of L-type Ca current (I_{CaL}) and PLB and RyR phosphorylation (Bers, 2002). Enhanced I_{CaL} leads to increased transmembrane ‘trigger’ Ca initiating the Ca-induced-Ca-release process, whereas phosphorylation of PLB and RyRs contributes to increased sarcoplasmic reticulum (SR) Ca uptake and release, respectively (Figure 1). Enhanced Ca and PLB phosphorylation favor faster relaxation via accelerated SR Ca reuptake. Phosphorylation of troponin I also contributes to positive lusitropy by accelerating dissociation of Ca from the myofilaments. This reduction in myofilament Ca sensitivity would, by itself, be expected to decrease contractility, but it is outweighed by the dramatic increase in intracellular Ca available for contraction (Bers, 2002).

The positive dromotropic effect includes increased SA nodal and atrioventricular (AV) nodal conduction velocity primarily mediated by increased I_{CaL} , which is a key component of the SA and AV nodal action potential upstroke (Bartos et al., 2015). Conduction velocity of the ventricular myocardium may also increase with β -AR activation (Wallace & Sarnoff, 1964;

Ng et al., 2007; Ajjola et al., 2017). PKA-mediated phosphorylation of I_{Na} may be involved (Herren et al., 2013) (see Section 3.2), but modulation of gap junctions is also a likely contributor (Figure 1, reviewed in (Campbell et al., 2014)). In the short-term, β -AR activation and increased cAMP impact phosphorylation and assembly of connexin43 (Cx43) (TenBroek et al., 2001; Somekawa et al., 2005), whereas more long-term effects may be due to adrenergically-mediated alterations in Cx43 turnover or expression (Salameh et al., 2006).

3.2 Arrhythmogenic mechanisms of β -adrenergic action: ionic bases

The complex cardiac electrophysiological and Ca handling consequences of sympathetic activation involve changes in transmembrane potential homeostasis via both direct influences on sarcolemmal ion channels and transporters as well as indirect changes in Ca signaling that acutely regulate transmembrane fluxes and can lead to remodeling in the chronic (pathologic) setting.

cAMP/PKA signaling modulates several ion currents including I_{Na} , I_{CaL} , I_K , as well as I_f in nodal cells (increased I_f expression in the working myocardium in disease may also result in ectopic activity that can be targeted by β -AR blockade). PKA-dependent phosphorylation of I_{Na} potentiates the current via both gating changes (Zhou et al., 2000) and by enhancing channel trafficking (Zhou et al., 2002). This may contribute to the sympathetically-mediated increase in conduction velocity and formation of reentrant arrhythmias after MI, which is often characterized by myocardial depolarization (Nattel et al., 2007). Late I_{Na} is also increased upon β -AR activation, mediated by both PKA and CaMKII (Wagner et al., 2006) increases (Hegyi et al., 2018). Thus, β -AR stimulation may enhance late I_{Na} , lengthen the AP, and increase the propensity to arrhythmogenic early afterdepolarizations (EADs), especially in disease states when late I_{Na} is enhanced (Clancy & Rudy, 1999).

I_{CaL} enhancement subsequent to PKA-dependent phosphorylation has been associated with prolonged AP and increased tendency for EADs, due to I_{CaL} reactivation during the AP plateau (phase 2 EADs) (Weiss et al., 2010). Increased Ca influx and consequent increase in Ca load, needed for positive inotropy, also favors spontaneous SR Ca release and Na/Ca exchange-mediated depolarization before or after completion of repolarization, leading to phase 3 EADs or delayed afterdepolarizations (DADs) (Bers, 2008), respectively. PKA-mediated phosphorylation of RyRs enhances their Ca sensitivity, which also favors DADs. Indeed, in genetically linked catecholaminergic polymorphic ventricular tachycardia (CPVT), DAD-induced arrhythmia is triggered by high sympathetic tone (for example during exercise) in patients with no myocardial damage (Laitinen et al., 2001).

K currents are also enhanced by cAMP/PKA signaling to counteract AP prolongation and limit Ca loading. Sympathetic stimulation modulates the delayed rectifier K current (Bartos et al., 2017). Under basal conditions the slowly activating I_{Ks} density is much lower than the rapidly activating component I_{Kr} in humans and other large mammals (Jost et al., 2007). However, β -AR stimulation increases I_{Ks} more than I_{Kr} (Banyasz et al., 2014) and counteracts I_{CaL} enhancement to prevent potentially harmful action potential duration (APD) prolongation. Indeed, exercise and stress are typical I_{CaL} arrhythmia triggers in congenital type 1 long QT syndrome (LQTS), linked to I_{Ks} loss of function (Schwartz et al., 2001), and can be prevented by β -AR blockade (Vincent et al., 2009). In a recent study, computer simulations

showed that increasing the I_{Ks}/I_{Kr} ratio, without changing the resulting APD, limits EAD occurrence in response to perturbations. This suggests the intriguing notion that I_{Ks} is more effective than I_{Kr} in stabilizing APD and suppressing EADs (Devenyi et al., 2017). The kinetic mismatch between faster phosphorylation-mediated activation of I_{CaL} and slower I_{Ks} increase upon β -AR activation transiently perturbs the balance of inward and outward currents (Liu et al., 2012). Computational modeling and simulations showed that this current imbalance that favors depolarization can prolong APD and favor EADs transiently (Xie et al., 2013).

At the tissue level, shorter APD and thus ERP can facilitate reentrant excitation, which is also promoted by structural changes such as fibrosis (Section 3.4) leading to slower impulse propagation. Spatial and temporal heterogeneity of PKA-dependent effects on depolarizing vs. repolarizing currents can also amplify dispersion of repolarization (DOR), thus increasing the likelihood for unidirectional conduction block and subsequent reentrant activity. Furthermore, faster I_{CaL} activation (vs. I_{Ks}) upon rapid β -AR stimuli transiently steepens APD restitution leading to spiral wave breakup and precipitating breakdown of ventricular tachycardia (VT) into ventricular fibrillation (VF) (Xie et al., 2014). Thus, the cellular triggering mechanisms discussed above combined with the tissue-level reentrant substrate can set the stage for β -AR arrhythmogenesis.

3.3 Arrhythmogenic mechanisms of β -adrenergic action: nerve remodeling in disease

The heart is extensively innervated (Pauza et al., 2002a; Pauza et al., 2002b), and recent detailed analysis of nerve-cardiomyocyte interaction has suggested that the sympathetic nerve-to-myocyte ratio is similar to the capillary-to-myocyte ratio (Freeman et al., 2014; Zaglia & Mongillo, 2017). Indeed, in the normal heart, it appears as if nearly every cardiomyocyte is in contact with one or more sympathetic nerves (Freeman et al., 2014; Zaglia & Mongillo, 2017). This density is not uniform, however, as there exist gradients in innervation from base to apex and from epi- to endocardium (Kawano et al., 2003). Importantly, cardiac sympathetic innervation undergoes extensive functional and anatomical remodeling during cardiovascular disease. Remodeling of sympathetic neurotransmission is arrhythmogenic and is associated with both hyper- and hypo-innervation as well as altered neurotransmitter content and release.

Regional hyper-innervation was one of the first identified forms of neural remodeling that was linked to arrhythmias in humans (Cao et al., 2000), and has now been well documented in various cardiac pathologies, including MI, HF, and AF (reviewed in (Chen et al., 2001)). The underlying mechanisms by which hyperinnervation leads to arrhythmia are likely due to excess NE release in a localized region of the heart, which exacerbates the cellular and tissue-level arrhythmogenic processes described above (Section 3.2). Indeed, our group demonstrated the mechanisms by which localized sympathetic stimulation leads to the generation of ventricular ectopic beats via synchronization of SR Ca overload and release (Myles et al., 2012). Excess catecholamines are also linked to downregulation of K currents and prolonged ventricular APD (Aflaki et al., 2014). Furthermore, both hyper-innervation and chronically elevated sympathetic drive present in cardiovascular disease can lead to decreased β -AR responsiveness and G-protein uncoupling (Soltysinska et al., 2011), which

may in turn lead to further elevations in sympathetic activity, thus perpetuating a vicious cycle of elevated sympathetic tone.

Many studies on the links between hyper-innervation and arrhythmia have focused on ventricular arrhythmias in MI or HF. However, hyper-innervation has also been documented in patients with chronic AF (Nguyen et al., 2009) and may be important in the initiation and maintenance of atrial tachyarrhythmias. Indeed, modulating autonomic function to reduce innervation or sympathetic activity has shown useful for AF control (reviewed in (Chen et al., 2014)), yet β -blockers are not typically used in AF rhythm control (see Section 4). Histological studies of the human pulmonary vein–left atrium junction showed that numerous autonomic nerves are present (Tan et al., 2006; Vaitkevicius et al., 2009), and that there is a mix of adrenergic and cholinergic fibers, suggesting that complex spatio-temporal interactions between sympathetic and parasympathetic activity may be involved in AF.

Paradoxically, sympathetic hypo-innervation (or denervation) is also linked to ventricular arrhythmias. Indeed, recent clinical studies have suggested that the degree of viable denervated myocardium is an independent predictor of ventricular arrhythmia risk and cardiac arrest (Boogers et al., 2010; Nishisato et al., 2010; Fallavollita et al., 2014). One explanation of these findings may be that any abnormal heterogeneity in sympathetic transmission is arrhythmogenic (Rubart & Zipes, 2005). Chronic hypo-innervation may also result in upregulation of β -ARs and adrenergic super-sensitivity of the myocardium, meaning that supra-physiological responses may occur with normal catecholamine exposure. Recent data from murine models suggests that the infarct remains devoid of sympathetic fibers following MI (Gardner & Habecker, 2013), and our group demonstrated that these denervated infarcts are in fact super-sensitive to adrenergic agonists, leading to electrophysiological heterogeneity and triggered activity (Gardner et al., 2015). Therefore, β -blockers may have significant anti-arrhythmic value even in denervated conditions.

3.4 Arrhythmogenic mechanisms of β -adrenergic action: hypertrophy and fibrosis

Cardiomyocyte hypertrophy and increased fibrosis are hallmarks of cardiovascular disease and both aspects of remodeling are associated with arrhythmias. Indeed, both organ enlargement and fibrotic remodeling create a vulnerable structural reentrant substrate, by generating longer conduction pathways for reentry, slowing conduction, and imposing unexcitable barriers that facilitate arrhythmia initiation and maintenance. Several studies have shown that β -AR agonists, including NE and isoproterenol, as well as β_1 -AR overexpression can produce cardiac hypertrophy and fibrosis *in vivo* (Engelhardt et al., 1999). Although myocyte hypertrophy may be an adaptive response to the increase in work load caused by myocardial β_1 -AR stimulation, there is evidence that direct adrenergic signaling may also be involved. In a seminal study in cultured cardiomyocytes, Simpson showed that NE-induced myocyte hypertrophy is mediated by α_1 -adrenergic receptors (Simpson, 1983). More recent work suggests that β_1 -AR signaling may also be involved. Pare and colleagues demonstrated that PKA phosphorylates a pool of perinuclear RyR2s, leading to increased local Ca, which in turn activates the pro-hypertrophic calcineurin-nuclear factor of activated T-cells transcription factor pathway (Pare et al., 2005). Cardiac non-myocytes, including fibroblasts also have adrenergic receptors, and stimulation of β_2 -

ARs in human and rodent cardiac fibroblasts leads to increased proliferation (Long et al, 1993; Turner et al, 2003). This raises the intriguing possibility that direct β -AR signaling in fibroblasts could be an important contributor to fibrosis in MI and HF and may explain some of the anti-fibrotic effects observed with β -blocker treatment.

4. Indications for clinical use of β -blockers

It is evident that β -blocker therapy may antagonize multiple direct and indirect arrhythmogenic effects of increased sympathetic activity. Depending on the arrhythmia type, β -blockers reduce proarrhythmic risk by preventing sympathetically-mediated triggers, functional reentrant substrates, and slowing of the SA- and AV-nodal rates. Table 1 lists currently used β -blockers, their mechanisms of action and therapeutic uses, including specific indications for arrhythmia.

β -blockers are a cornerstone of anti-arrhythmic drug therapy. β -blockers are generally safe agents that effectively suppress ventricular ectopic beats and arrhythmia, and prevent sudden cardiac death in a wide array of cardiac diseases (Al-Khatib et al., 2018). According to guidelines, β -blockers are indicated in all patients, except those with AV block, bradycardia, or asthma, and recommended in all HF patients regardless of baseline rhythm, β -blockers are also used for control of ventricular rates to avoid rapid irregular ventricular activation due to rapid and irregular atrial firing during AF (January et al., 2014).

AF.

β -blockers are first line therapy for ventricular rate control in AF (January et al., 2014). They act by slowing conduction through the AV node, have been proven superior in ventricular rate control especially with exercise, and are preferred to digoxin and Ca channel blockers in patients with MI or HF. β -blockers may be avoided in patients with chronic pulmonary disease and at risk of bronchoconstriction. In acute AF settings, intravenous administration of esmolol, propranolol, and metoprolol has been shown effective; in chronic AF, oral administration of β -blockers, including atenolol, bisoprolol, metoprolol, nadolol, propranolol, and sotalol (a K channel blocker), is effective for ventricular rate control (January et al., 2014). Of note, comparison of different β -blockers demonstrated that carvedilol is less effective than metoprolol for rate control (Vittorio et al., 2008).

β -blockers have a weak antiarrhythmic action compared to Class I (Na channel blockers) and Class III agents (K channel blockers), and are not generally considered as atrial rhythm control agents (January et al., 2014). However, β -blockers may be beneficial in some patients in combination with an antiarrhythmic drug. Note that Class III amiodarone, the most effective rhythm control agent in patients with AF, is also a β -AR antagonist. β -blockers may be helpful for AF prevention in patients with adrenergically-mediated AF, for example linked to stress or anxiety, and in patients following cardiothoracic surgery, with likely elevated postoperative sympathetic tone. On the other hand, they could be detrimental in vagally-mediated AF. Further, β -blockers have been shown to prevent the occurrence of AF in patients with systolic HF (Nasr et al., 2007).

MI.

β -blockers are known to decrease mortality both during acute MI and with long-term administration following MI. Many randomized clinical studies of β -blockers were performed prior to routine anti-platelet and statin therapy, so the absolute benefit may be lower, but initial clinical trials indicated a 10-25% reduction in mortality in patients treated with timolol, metoprolol, atenolol, or propranolol (Norwegian Multicenter Study, 1981; Hjalmarson et al., 1983; 1986; Chadda et al., 1986). Current recommendations for acute MI include cardioselective oral β -blockers, such as metoprolol or atenolol. For long-term administration after MI, agents that lack sympathomimetic activity are preferred (Antman et al., 2004; Antman et al., 2008).

β -blockade effects include (i) decreased myocardial oxygen demand and reduction of ischemic burden, due to lowering of heart rate, myocardial contractility, and blood pressure; (ii) prevention of maladaptive ventricular remodeling and failure, and (iii) decreased risk of VF and sudden cardiac death, as demonstrated in both experimental and clinical studies. Specific anti-arrhythmic effects may include lengthening of the ventricular effective refractory period, suppression of triggered activity and automaticity, attenuation of electrophysiological heterogeneity (e.g., caused by MI-induced hypo- or hyper-innervation), and slowing of heart rate. Recent experimental evidence also suggests that sympathetic nerve activity can modulate conduction through putative reentrant circuits in the infarct border zone, making them more prone to conduction block (Ajjola et al., 2017), suggesting another possible anti-arrhythmic mechanism of β -blockade following MI.

HF.

β -blockers are a mainstay of HF therapy (Yancy et al., 2013; Yancy et al., 2017). Three β -blockers, carvedilol (Packer et al., 2001), metoprolol (1999b), and bisoprolol (1999a), have been studied in clinical trials, whereby chronic treatment has been demonstrated to improve symptoms, reduce hospitalization, and enhance survival when used in addition to diuretics and angiotensin converting enzyme (ACE) inhibitors. β -blockers are only contraindicated in acute decompensated setting, whereby the negative inotropy is detrimental. β -blockers prevent sudden cardiac death in patients with systolic HF and reduce all-cause mortality, i.e., adverse effects of catecholamine stimulation, including increases in heart rate and myocardial energy requirements, maladaptive remodeling due to cell hypertrophy and death, fibrosis, proarrhythmia, and inappropriate stimulation of other pathways such as the renin-angiotensin-aldosterone system.

HF-induced remodeling involves key nodes of the cAMP/PKA signaling cascade, including downregulation of β_1 - (~60%) and upregulation in β_2 - (~40%) and β_3 -AR, switching from G_s to G_i coupling, and activation of G protein-independent pathways, as recently reviewed (de Lucia et al., 2018). Extensive ionic remodeling also occurs (Nattel et al., 2007; Bartos et al., 2015) and involves downregulation of several K channels, i.e., those carrying I_{Kr} , I_{Ks} , I_{K1} , and I_{to} and consequent AP prolongation, increased risk for EADs, and increased DOR and dispersion of refractoriness. Ca handling abnormalities in HF include increased spontaneous Ca release and risk for DADs (Bers, 2006). Alterations in gap junction and structural remodeling (Burchfield et al., 2013), involving myocyte hypertrophy, organ

dilation, and fibrosis, contribute to slowing of conduction that can lead to unidirectional conduction block and predispose to reentry. Thus, the antiarrhythmic action of β -blockers in HF might be mediated by attenuating both the triggers and the functional or structural arrhythmia substrates.

LQTS.

β -blockers are a mainstay of treatment for LQTS (Ackerman et al., 2017). β -blockers are recommended in patients diagnosed with LQTS, and should be considered in patients that carry a causative LQTS mutation but have normal QT interval. Increased sympathetic tone (e.g., during exercise) is one of the most important arrhythmia triggers in LQT1 (which is caused by mutations in the *KCNQ1* gene leading to reduction in I_{Ks} current), and can be prevented by β -blockers (Schwartz et al., 2001; Vincent et al., 2009). In LQT2 (caused by loss of function of I_{Kr}), β -blockers are thought to be less effective than in LQT1. Recent studies comparing efficacy of different β -blockers, reviewed in (Ackerman et al., 2017), showed that propranolol and nadolol were similarly effective, whereas metoprolol had significantly less anti-arrhythmic efficacy. Importantly, when comparing the efficacy of different β -blocking agents independently for LQT1 and for LQT2, nadolol had the greatest efficacy among the more severely affected LQT2 patient group. It has been shown that β -blockers reduce risk in LQT3 patients (Wilde et al., 2016), despite prior studies indicating that β -blockers were not as effective in LQT3 as compared to LQT1 or LQT2.

CPVT.

CPVT is caused by defective inter-domain interaction within the RyR2 (Yamamoto et al., 2000), which enhances arrhythmias by promoting diastolic Ca leak. First-line treatment for CPVT patients includes exercise restriction and β -blockade (with agents lacking intrinsic sympathomimetic activity) (Ackerman et al., 2017). According to guidelines, β -blockers are recommended in all CPVT patients with documented ventricular arrhythmias (either spontaneous or stress-induced), and should be considered for asymptomatic mutation carriers even after a negative exercise stress test. Left cardiac sympathetic denervation might be an option for CPVT patients that are intolerant to β -blockers, but its efficacy remains to be fully assessed. Preliminary data on a small group of CPVT patients suggest that flecainide (a Na channel blocker known to interact with RyRs) significantly reduces arrhythmias and should be considered in combination with β -blockers when arrhythmia control is incomplete (Kannankeril et al., 2017). An implantable cardioverter-defibrillator (ICD) is indicated in patients that do not respond to β -blockade and flecainide (Ackerman et al., 2017).

5. Key future work

While available experimental and clinical evidence supports an important role of β -AR antagonism in counteracting acute and chronic detrimental effects of β -AR stimulation, important key questions remain to be addressed, that could pave the way for major new developments in β -blockade strategies and their therapeutic uses in cardiac disease and arrhythmia.

Exploiting drugs' receptor-specific β -AR agonism and antagonism.

While acute activation of β_1 - and β_2 -ARs exerts positive tropic actions, chronic activation of β_1 -ARs causes (mal)adaptive effects including cardiac hypertrophy and fibrosis, and cell death, which contribute to the development of HF, and lethal arrhythmias. On the other hand, long term stimulation of β_2 -ARs improves myocyte survival and overall cardiac function (Xiao, 2001). Further, while the functional significance of β_3 -ARs is incompletely understood, preclinical studies showed that β_3 -ARs can activate different signaling pathways that can protect the heart. For example, stimulation of β_3 -ARs activates a downstream NO-GC-cGMP pathway that limits Ca influx and is thought to be cardioprotective (Cannavo & Koch, 2017). Thus, sustained activation of β_2 - and β_3 -ARs combined with β_1 -AR blockade could be a new receptor-specific therapeutic approach for the chronic HF treatment. In addition to the cardioprotective effects of β_2 - and β_3 -AR stimulation, activation of β_2 -AR stimulation has direct vasodilatory effects, and β_3 -AR activation has been shown to increase lipolysis and may also have antidepressant activity (Ferrer-Lorente et al., 2005; Consoli et al., 2007). Thus, such antagonist/agonist drugs could have myriad positive effects in patients with cardiovascular disease (see Table 1).

Targeting compartmentalized signaling.

There is solid evidence that activation of β -ARs can generate spatially restricted pools of cAMP that in turn lead to localized intracellular (rather than global) PKA activation and result in specific downstream functional effects (Surdo et al., 2017). For example, the intensity, duration, and spatial range of cAMP signals is strongly modulated by cAMP-degrading phosphodiesterase activity and localization (Zaccolo & Pozzan, 2002; Surdo et al., 2017). Thus, using drugs that target specific cAMP pools, rather than affecting global intracellular cAMP levels, could be a promising strategy to improve therapeutic specificity (Zaccolo, 2009). This requires detailed understanding of the spatial organization, regulation and functional role of cAMP compartments. Furthermore, even in the presence of uniform cAMP signals, distinct domains of PKA-phosphorylated proteins can be obtained due to subcellular heterogeneity in protein phosphatase distribution (Burdyga et al., 2018), leading to differential phosphorylation of various downstream PKA targets that promote specific cardiac responses. For example, activation of protein phosphatase 1 in human HF opposes increased kinase activity and attenuates arrhythmogenic Ca leak (Fischer et al., 2018). Real-time imaging of cAMP and resulting PKA activity using FRET-based sensors has greatly contributed to our understanding of compartmentalized cAMP signaling (Zaccolo & Pozzan, 2002; Surdo et al., 2017). In particular, novel sensors targeted to protein complexes involved in excitation-contraction coupling have begun to address crucial questions regarding the size and spatial distribution of distinct cAMP compartments, the magnitude and kinetics of cAMP signals within each compartment, and the specific role of individual compartments in regulating cell function (Nikolaev et al., 2010; Surdo et al., 2017). For example, β_2 -ARs were shown to be concentrated in the transverse tubules, leading to localized cAMP signal in healthy cells. In HF, however, these receptors were redistributed to the cell crest, leading to diffuse receptor-mediated cAMP signaling (Nikolaev et al., 2010). These approaches have provided original insight into the regulation of cardiac excitation and contraction in health and disease with profound implications for therapy (Surdo et al., 2017).

Evaluating the interaction between β -AR and other signaling pathways.

A clear link has been established between enhanced sympathetic activation and ventricular arrhythmias in both animal models and humans with cardiac disease. Excessive β -AR activation is well documented in HF, but the complex and multifaceted nature of the disease suggests that multiple other signaling pathways are perturbed. Notably, many of them might crosstalk with the β -AR system. For example, the renin-angiotensin-aldosterone system (RAAS) is also chronically active in HF. Experimental evidence indicates both that β -AR blockade may diminish activity of the RAAS, and that targeting the RAAS may reduce sympathetic nerve activity (Goldsmith, 2004), suggesting that combination therapy that suppresses each individual system involves a virtuous (negative feedback) cycle.

Stimulation of various signaling pathways, altered metabolism, and increased oxidative stress and their complex interactions may exert electrophysiological abnormalities acutely, and accumulation of these changes (e.g., in chronic pathological settings) may cause prolonged alterations in cardiac signal transduction and gene expression. In cardiac myocytes, β -adrenergic stimulation, via both direct (EPAC, NO)(Pereira et al., 2017) and indirect (increased Ca) mechanisms, enhances the activity of the Ca/calmodulin-dependent protein kinase (CaMKII), which is overexpressed and hyper-activated in HF, and critically regulates cellular subsystems participating in acute mechanical and electrical abnormalities in HF and models of adrenergic stimulation as well as long term cardiac remodeling in HF (Grandi & Dobrev, 2018). CaMKII phosphorylates a number of downstream targets that play important roles in excitation-contraction coupling (Figure 1), and many of these proteins are also targets of PKA-dependent phosphorylation. The relative contribution of these kinases to proarrhythmic functional alterations has begun to be defined. For example, it has been shown that while both kinases are involved in RyR dysregulation in human hypertrophy, in end stage human HF, CaMKII predominates to induce arrhythmogenic Ca leak (Fischer et al., 2013). Recent data also revealed differential modulations of I_{NaL} by PKA and CaMKII at different phases of the action potential (Hegyi et al., 2018). It has been hypothesized that in HF, synergy between CaMKII upregulation and altered Na and Ca fluxes can lead to a vicious (positive feedback) cycle perpetuating the arrhythmia, which is further accentuated during β -adrenergic stimulation. Thus, β -AR blockade could counteract arrhythmias via both direct effects and by de-escalating the synergistic interaction between CaMKII and β -AR signaling (Bers, 2005).

Understanding sex-based differences.

There are well-known sex differences in male and female cardiac electrophysiological properties (Ambrosi et al., 2013). For example, women have a prolonged baseline APD and QT interval, which lead to increased risk for drug-induced torsades de pointe (TdP) in females (Pham et al., 2001; Salama & Bett, 2014; Kurokawa et al., 2016). Moreover, the mechanisms of arrhythmias in HF may also differ between males and females, with cells from male failing hearts demonstrating an increase in Ca leak, spark frequency, and triggered activity compared to failing female cells (Fischer et al., 2016). Interestingly, normal male rabbit hearts also have a stronger, and more arrhythmogenic response to β -AR stimulation, showing an increase in triggered activity, despite similar increases in diastolic Ca leak (Hoeker et al., 2014). Further, heart rate variability studies suggest underlying sex

differences in autonomic control of the cardiovascular system, whereby women have higher degrees of parasympathetic activation, whereas sympathetic-mediated responses predominate in men (Pothineni et al., 2016). This has been associated with an increased propensity in women of AF due to extensive vagal innervation of the atrial muscle sleeves extending into the pulmonary veins. Furthermore, the ORBIT-AF registry revealed lower rates of use of β -blockers as rate-control agents in women (as opposed to digoxin) (Piccini et al., 2016), though the reason is unknown. These observations also suggest that there may be significant sex differences in efficacy of β -blocker therapy for arrhythmia prevention. Interestingly, in the MERIT-HF study, which assessed the efficacy of metoprolol, the 23% of women included were the only subgroup in which no favorable effect on mortality was observed (although the women in this study were still 37% less likely to die than men) (Group, 1999). Yet, post-hoc analyses of MERIT-HF and other studies do not show sex differences in mortality with β -blocker therapy for HF (Ghali et al., 2002). Importantly, while these data regarding mortality do not indicate whether these deaths were arrhythmic in nature, the mixed observations suggest an urgent need to better understand the mechanisms of underlying sex differences, which may suggest more personalized approaches to β -blocker therapy.

6. Conclusions

The effects of β -adrenergic stimulation on cardiac electrical activity and remodeling involve structural and functional changes occurring over multiple time- and spatial-scales. While significant progress has been made towards understanding the role of β -AR signaling in heart disease and arrhythmias, a comprehensive quantitative and functional understanding of the role of autonomic stimulation in normal cardiac electrophysiology and life-threatening arrhythmias is still lacking. We contend that defining the structural and functional anatomy of cardiac innervation, and linking neural structure and function to multiscale cardiac electrophysiology, e.g., via computational modeling approaches and simulation (Morotti & Grandi, 2018), is a necessary step to improve our understanding of this complex system. Integrating these data from the sub-cellular, tissue, and multi-organ levels could accelerate our understanding of the complex network of pathways involved, predict mechanisms underlying the interaction between adrenergic activation and the functional cardiac substrate, and facilitate identification and specific targeting of arrhythmia provoking conditions by autonomic drugs.

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References

- (1986). Randomised trial of intravenous atenolol among 16 027 cases of suspected acute myocardial infarction: ISIS-1. First International Study of Infarct Survival Collaborative Group. *Lancet* 2, 57–66. [PubMed: 2873379]

- (1999a). The Cardiac Insufficiency Bisoprolol Study II (CIBIS-II): a randomised trial. *Lancet* 353, 9–13. [PubMed: 10023943]
- (1999b). Effect of metoprolol CR/XL in chronic heart failure: Metoprolol CR/XL Randomised Intervention Trial in Congestive Heart Failure (MERIT-HF). *Lancet* 353, 2001–2007. [PubMed: 10376614]
- Ackerman MJ, Priori SG, Dubin AM, Kowey P, Linker NJ, Slotwiner D, Triedman J, Van Hare GF & Gold MR. (2017). Beta-blocker therapy for long QT syndrome and catecholaminergic polymorphic ventricular tachycardia: Are all beta-blockers equivalent? *Heart Rhythm* 14, e41–e44. [PubMed: 27659101]
- Aflaki M, Qi XY, Xiao L, Ordog B, Tadevosyan A, Luo X, Maguy A, Shi Y, Tardif JC & Nattel S. (2014). Exchange protein directly activated by cAMP mediates slow delayed-rectifier current remodeling by sustained beta-adrenergic activation in guinea pig hearts. *Circ Res* 114, 993–1003. [PubMed: 24508724]
- Ajjola OA, Lux RL, Khahera A, Kwon O, Aliotta E, Ennis DB, Fishbein MC, Ardell JL & Shivkumar K. (2017). Sympathetic modulation of electrical activation in normal and infarcted myocardium: implications for arrhythmogenesis. *Am J Physiol Heart Circ Physiol* 312, H608–H621. [PubMed: 28087519]
- Al-Khatib SM, Stevenson WG, Ackerman MJ, Bryant WJ, Callans DJ, Curtis AB, Deal BJ, Dickfeld T, Field ME, Fonarow GC, Gillis AM, Granger CB, Hammill SC, Hlatky MA, Joglar JA, Kay GN, Matlock DD, Myerburg RJ & Page RL. (2018). 2017 AHA/ACC/HRS Guideline for Management of Patients With Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death: Executive Summary. *Circulation* 138, e210–e271. [PubMed: 29084733]
- Ambrosi CM, Yamada KA, Nerbonne JM & Efimov IR. (2013). Gender differences in electrophysiological gene expression in failing and non-failing human hearts. *PLoS One* 8, e54635. [PubMed: 23355885]
- Antman EM, Anbe DT, Armstrong PW, Bates ER, Green LA, Hand M, Hochman JS, Krumholz HM, Kushner FG, Lamas GA, Mullany CJ, Ornato JP, Pearle DL, Sloan MA, Smith SC Jr., Alpert JS, Anderson JL, Faxon DP, Fuster V, Gibbons RJ, Gregoratos G, Halperin JL, Hiratzka LF, Hunt SA, Jacobs AK & Ornato JP. (2004). ACC/AHA guidelines for the management of patients with ST-elevation myocardial infarction; A report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Committee to Revise the 1999 Guidelines for the Management of patients with acute myocardial infarction). *J Am Coll Cardiol* 44, E1–E211. [PubMed: 15358047]
- Antman EM, Hand M, Armstrong PW, Bates ER, Green LA, Halasyamani LK, Hochman JS, Krumholz HM, Lamas GA, Mullany CJ, Pearle DL, Sloan MA, Smith Sc Jr., Writing Committee M, Anbe DT, Kushner fG, Ornato JP, Jacobs AK, Adams CD, Anderson JL, Buller CE, Creager MA, Ettinger SM, Halperin JL, Hunt SA, Lytle BW, Nishimura R, Page RL, Riegel B, Tarkington LG & Yancy CW. (2008). 2007 Focused Update of the ACC/AHA 2004 Guidelines for the Management of Patients With ST-Elevation Myocardial Infarction: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines: developed in collaboration With the Canadian Cardiovascular Society endorsed by the American Academy of Family Physicians: 2007 Writing Group to Review New Evidence and Update the ACC/AHA 2004 Guidelines for the Management of Patients With ST-Elevation Myocardial Infarction, Writing on Behalf of the 2004 Writing Committee. *Circulation* 117, 296–329. [PubMed: 18071078]
- Banyasz T, Jian Z, Horvath B, Khabbaz S, Izu LT & Chen-Izu Y. (2014). Beta-adrenergic stimulation reverses the I Kr-I Ks dominant pattern during cardiac action potential. *Pflugers Arch* 466, 2067–2076. [PubMed: 24535581]
- Bartos DC, Grandi E & Ripplinger CM. (2015). Ion Channels in the Heart. *Compr Physiol* 5, 1423–1464. [PubMed: 26140724]
- Bartos DC, Morotti S, Ginsburg KS, Grandi E & Bers DM. (2017). Quantitative analysis of the Ca(2+) - dependent regulation of delayed rectifier K(+) current IKs in rabbit ventricular myocytes. *J Physiol* 595, 2253–2268. [PubMed: 28008618]
- Bers DM. (2002). Cardiac excitation-contraction coupling. *Nature* 415, 198–205. [PubMed: 11805843]
- Bers DM. (2005). Beyond beta blockers. *Nat Med* 11, 379–380. [PubMed: 15812516]

- Bers DM. (2006). Altered cardiac myocyte Ca regulation in heart failure. *Physiology (Bethesda)* 21, 380–387. [PubMed: 17119150]
- Bers DM. (2008). Calcium cycling and signaling in cardiac myocytes. *Annu Rev Physiol* 70, 23–49. [PubMed: 17988210]
- Boogers MJ, Borleffs CJ, Henneman MM, van Bommel RJ, van Ramshorst J, Boersma E, Dibbets-Schneider P, Stokkel MP, van der Wall EE, Schalij MJ & Bax JJ. (2010). Cardiac sympathetic denervation assessed with 123-iodine metaiodobenzylguanidine imaging predicts ventricular arrhythmias in implantable cardioverter-defibrillator patients. *J Am Coll Cardiol* 55, 2769–2777. [PubMed: 20538172]
- Bristow MR, Ginsburg R, Umans V, Fowler M, Minobe W, Rasmussen R, Zera P, Menlove R, Shah P, Jamieson S & et al. (1986). Beta 1- and beta 2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta 1-receptor down-regulation in heart failure. *Circ Res* 59, 297–309. [PubMed: 2876788]
- Brunton LL, Hilal-Dandan R & Knollmann BC. (2018). *Goodman & Gilman's: The Pharmacological Basis of Therapeutics*, 13e. McGraw-Hill Education.
- Burchfield JS, Xie M & Hill JA. (2013). Pathological ventricular remodeling: mechanisms: part 1 of 2. *Circulation* 128, 388–400. [PubMed: 23877061]
- Burdyga A, Surdo NC, Monterisi S, Di Benedetto G, Grisan F, Penna E, Pellegrini L, Zaccolo M, Bortolozzi M, Swietach P, Pozzan T & Lefkimmatis K. (2018). Phosphatases control PKA-dependent functional microdomains at the outer mitochondrial membrane. *Proc Natl Acad Sci U S A* 115, E6497–E6506. [PubMed: 29941564]
- Campbell AS, Johnstone SR, Baillie GS & Smith G. (2014). beta-Adrenergic modulation of myocardial conduction velocity: Connexins vs. sodium current. *J Mol Cell Cardiol* 77, 147–154. [PubMed: 25453599]
- Cannavo A & Koch WJ. (2017). Targeting beta3-Adrenergic Receptors in the Heart: Selective Agonism and beta-Blockade. *J Cardiovasc Pharmacol* 69, 71–78. [PubMed: 28170359]
- Cannon WB. (1915). Bodily changes in pain, hunger, fear and rage, an account of recent researches into the function of emotional excitement. New York and London, D. Appleton and Co.
- Cao JM, Fishbein MC, Han JB, Lai WW, Lai AC, Wu TJ, Czer L, Wolf PL, Denton TA, Shintaku IP, Chen PS & Chen LS. (2000). Relationship between regional cardiac hyperinnervation and ventricular arrhythmia. *Circulation* 101, 1960–1969. [PubMed: 10779463]
- Chadda K, Goldstein S, Byington R & Curb JD. (1986). Effect of propranolol after acute myocardial infarction in patients with congestive heart failure. *Circulation* 73, 503–510. [PubMed: 3948357]
- Chen PS, Chen LS, Cao JM, Sharifi B, Karagueuzian HS & Fishbein MC. (2001). Sympathetic nerve sprouting, electrical remodeling and the mechanisms of sudden cardiac death. *Cardiovasc Res* 50, 409–416. [PubMed: 11334845]
- Chen PS, Chen LS, Fishbein MC, Lin SF & Nattel S. (2014). Role of the autonomic nervous system in atrial fibrillation: pathophysiology and therapy. *Circ Res* 114, 1500–1515. [PubMed: 24763467]
- Clancy CE & Rudy Y. (1999). Linking a genetic defect to its cellular phenotype in a cardiac arrhythmia. *Nature* 400, 566–569. [PubMed: 10448858]
- Consoli D, Leggio GM, Mazzola C, Micale V & Drago F. (2007). Behavioral effects of the beta3 adrenoceptor agonist SR58611A: is it the putative prototype of a new class of antidepressant/anxiolytic drugs? *Eur J Pharmacol* 573, 139–147. [PubMed: 17669397]
- de Lucia C, Eguchi A & Koch WJ. (2018). New Insights in Cardiac beta-Adrenergic Signaling During Heart Failure and Aging. *Front Pharmacol* 9, 904. [PubMed: 30147654]
- Devenyi RA, Ortega FA, Groenendaal W, Krogh-Madsen T, Christini DJ & Sobie EA. (2017). Differential roles of two delayed rectifier potassium currents in regulation of ventricular action potential duration and arrhythmia susceptibility. *J Physiol* 595, 2301–2317. [PubMed: 27779762]
- DiFrancesco D & Tortora P. (1991). Direct activation of cardiac pacemaker channels by intracellular cyclic AMP. *Nature* 351, 145–147. [PubMed: 1709448]
- Engelhardt S, Hein L, Wiesmann F & Lohse MJ. (1999). Progressive hypertrophy and heart failure in beta1-adrenergic receptor transgenic mice. *Proc Natl Acad Sci U S A* 96, 7059–7064. [PubMed: 10359838]

- Fallavollita JA, Heavey BM, Luisi AJ Jr., Michalek SM, Baldwa S, Mashtare TL Jr., Hutson AD, Dekemp RA, Haka MS, Sajjad M, Cimato TR, Curtis AB, Cain ME & Canty Jm Jr. (2014). Regional myocardial sympathetic denervation predicts the risk of sudden cardiac arrest in ischemic cardiomyopathy. *J Am Coll Cardiol* 63, 141–149. [PubMed: 24076296]
- Ferrer-Lorente R, Cabot C, Fernandez-Lopez JA & Alemany M. (2005). Combined effects of oleoyl-estrone and a beta3-adrenergic agonist (CL316,243) on lipid stores of diet-induced overweight male Wistar rats. *Life Sci* 77, 2051–2058. [PubMed: 15935402]
- Fischer TH, Eiringhaus J, Dybkova N, Saadatmand A, Pabel S, Weber S, Wang Y, Kohn M, Tirilomis T, Ljubojevic S, Renner A, Gummert J, Maier LS, Hasenfuss G, El-Armouche A & Sossalla S. (2018). Activation of protein phosphatase 1 by a selective phosphatase disrupting peptide reduces sarcoplasmic reticulum Ca(2+) leak in human heart failure. *Eur J Heart Fail* 20, 1673–1685. [PubMed: 30191648]
- Fischer TH, Herting J, Eiringhaus J, Pabel S, Hartmann NH, Ellenberger D, Friedrich M, Renner A, Gummert J, Maier LS, Zabel M, Hasenfuss G & Sossalla S. (2016). Sex-dependent alterations of Ca2+ cycling in human cardiac hypertrophy and heart failure. *Europace* 18, 1440–1448. [PubMed: 26493982]
- Fischer TH, Herting J, Tirilomis T, Renner A, Neef S, Toischer K, Ellenberger D, Forster A, Schmitt JD, Gummert J, Schondube FA, Hasenfuss G, Maier LS & Sossalla S. (2013). Ca2+/calmodulin-dependent protein kinase II and protein kinase A differentially regulate sarcoplasmic reticulum Ca2+ leak in human cardiac pathology. *Circulation* 128, 970–981. [PubMed: 23877259]
- Freeman K, Tao W, Sun H, Soonpaa MH & Rubart M. (2014). In situ three-dimensional reconstruction of mouse heart sympathetic innervation by two-photon excitation fluorescence imaging. *J Neurosci Methods* 221, 48–61. [PubMed: 24056230]
- Gardner RT & Habecker BA. (2013). Infarct-derived chondroitin sulfate proteoglycans prevent sympathetic reinnervation after cardiac ischemia-reperfusion injury. *J Neurosci* 33, 7175–7183. [PubMed: 23616527]
- Gardner RT, Wang L, Lang BT, Cregg JM, Dunbar CL, Woodward WR, Silver J, Ripplinger CM & Habecker BA. (2015). Targeting protein tyrosine phosphatase sigma after myocardial infarction restores cardiac sympathetic innervation and prevents arrhythmias. *Nat Commun* 6, 6235. [PubMed: 25639594]
- Ghali JK, Pina IL, Gottlieb SS, Deedwania PC, Wikstrand JC & Group M-HS. (2002). Metoprolol CR/XL in female patients with heart failure: analysis of the experience in Metoprolol Extended-Release Randomized Intervention Trial in Heart Failure (MERIT-HF). *Circulation* 105, 1585–1591. [PubMed: 11927527]
- Goldsmith SR. (2004). Interactions between the sympathetic nervous system and the RAAS in heart failure. *Curr Heart Fail Rep* 1, 45–50. [PubMed: 16036024]
- Grandi E & Dobrev D. (2018). Non-ion channel therapeutics for heart failure and atrial fibrillation: Are CaMKII inhibitors ready for clinical use? *J Mol Cell Cardiol* 121, 300–303. [PubMed: 29079077]
- Grandi E, Sanguinetti MC, Bartos DC, Bers DM, Chen-Izu Y, Chiamvimonvat N, Colecraft HM, Delisle BP, Heijman J, Navedo MF, Noskov S, Proenza C, Vandenberg JJ & Yarov-Yarovoy V. (2017). Potassium channels in the heart: structure, function and regulation. *J Physiol* 595, 2209–2228. [PubMed: 27861921]
- Group M-HS. (1999). Effect of metoprolol CR/XL in chronic heart failure: Metoprolol CR/XL Randomised Intervention Trial in Congestive Heart Failure (MERIT-HF). *Lancet* 353, 2001–2007. [PubMed: 10376614]
- Hegyi B, Banyasz T, Izu LT, Belardinelli L, Bers DM & Chen-Izu Y. (2018). beta-adrenergic regulation of late Na(+) current during cardiac action potential is mediated by both PKA and CaMKII. *J Mol Cell Cardiol* 123, 168–179. [PubMed: 30240676]
- Herren AW, Bers DM & Grandi E. (2013). Post-translational modifications of the cardiac Na channel: contribution of CaMKII-dependent phosphorylation to acquired arrhythmias. *Am J Physiol Heart Circ Physiol* 305, H431–445. [PubMed: 23771687]
- Hjalmarson A, Herlitz J, Holmberg S, Ryden L, Swedberg K, Vedin A, Waagstein F, Waldenström A, Waldenström J, Wedel H, Wilhelmsson L & Wilhelmsson C. (1983). The Goteborg metoprolol trial.

Effects on mortality and morbidity in acute myocardial infarction. *Circulation* 67, 126–32. [PubMed: 6342837]

Hoeker GS, Hood AR, Katra RP, Poelzing S & Pogwizd SM. (2014). Sex differences in beta-adrenergic responsiveness of action potentials and intracellular calcium handling in isolated rabbit hearts. *PLoS One* 9, e111411. [PubMed: 25340795]

January CT, Wann LS, Alpert JS, Calkins H, Cigarroa JE, Cleveland JC Jr., Conti JB, Ellinor PT, Ezekowitz MD, Field ME, Murray KT, Sacco RL, Stevenson WG, Tchou PJ, Tracy CM, Yancy CW & Members AATF. (2014). 2014 AHA/ACC/HRS guideline for the management of patients with atrial fibrillation: a report of the American College of Cardiology/American Heart Association Task Force on practice guidelines and the Heart Rhythm Society. *Circulation* 130, e199–267. [PubMed: 24682347]

Jost N, Papp JG & Varro A. (2007). Slow delayed rectifier potassium current (IKs) and the repolarization reserve. *Ann Noninvasive Electrocardiol* 12, 64–78. [PubMed: 17286653]

Kannankeril PJ, Moore JP, Cerrone M, Priori SG, Kertesz NJ, Ro PS, Batra AS, Kaufman ES, Fairbrother DL, Saarel EV, Etheridge SP, Kanter RJ, Carboni MP, Dzurik MV, Fountain D, Chen H, Ely EW, Roden DM & Knollmann BC. (2017). Efficacy of Flecainide in the Treatment of Catecholaminergic Polymorphic Ventricular Tachycardia: A Randomized Clinical Trial. *JAMA Cardiol* 2, 759–766. [PubMed: 28492868]

Kawano H, Okada R & Yano K. (2003). Histological study on the distribution of autonomic nerves in the human heart. *Heart Vessels* 18, 32–39. [PubMed: 12644879]

Kurokawa J, Kodama M, Clancy CE & Furukawa T. (2016). Sex hormonal regulation of cardiac ion channels in drug-induced QT syndromes. *Pharmacol Ther* 168, 23–28. [PubMed: 27595633]

Laitinen PJ, Brown KM, Piippo K, Swan H, Devaney JM, Brahmabhatt B, Donarum EA, Marino M, Tiso N, Viitasalo M, Toivonen L, Stephan DA & Kontula K. (2001). Mutations of the cardiac ryanodine receptor (RyR2) gene in familial polymorphic ventricular tachycardia. *Circulation* 103, 485–490. [PubMed: 11157710]

Lakatta EG & DiFrancesco D. (2009). What keeps us ticking: a funny current, a calcium clock, or both? *J Mol Cell Cardiol* 47, 157–170. [PubMed: 19361514]

Liu GX, Choi BR, Ziv O, Li W, de Lange E, Qu Z & Koren G. (2012). Differential conditions for early after-depolarizations and triggered activity in cardiomyocytes derived from transgenic LQT1 and LQT2 rabbits. *J Physiol* 590, 1171–1180. [PubMed: 22183728]

Long CS, Hartogensis WE & Simpson PC. (1993). Beta-adrenergic stimulation of cardiac non-myocytes augments the growth-promoting activity of non-myocyte conditioned medium. *J Mol Cell Cardiol* 25, 915–925. [PubMed: 7505339]

Morotti S & Grandi E. (2018). Quantitative systems models illuminate arrhythmia mechanisms in heart failure: Role of the Na(+) -Ca(2+) -Ca(2+) /calmodulin-dependent protein kinase II-reactive oxygen species feedback. *Wiley Interdiscip Rev Syst Biol Med*, e1434. [PubMed: 30015404]

Myles RC, Wang L, Kang C, Bers DM & Ripplinger CM. (2012). Local beta-adrenergic stimulation overcomes source-sink mismatch to generate focal arrhythmia. *Circ Res* 110, 1454–1464. [PubMed: 22539768]

Nasr IA, Bouzamondo A, Hulot JS, Dubourg O, Le Heuzey JY & Lechat P. (2007). Prevention of atrial fibrillation onset by beta-blocker treatment in heart failure: a meta-analysis. *Eur Heart J* 28, 457–462. [PubMed: 17289748]

Nattel S, Maguy A, Le Bouter S & Yeh YH. (2007). Arrhythmogenic ion-channel remodeling in the heart: heart failure, myocardial infarction, and atrial fibrillation. *Physiol Rev* 87, 425–456. [PubMed: 17429037]

Ng GA, Brack KE, Patel VH & Coote JH. (2007). Autonomic modulation of electrical restitution, alternans and ventricular fibrillation initiation in the isolated heart. *Cardiovasc Res* 73, 750–760. [PubMed: 17217937]

Nguyen BL, Fishbein MC, Chen LS, Chen PS & Masroor S. (2009). Histopathological substrate for chronic atrial fibrillation in humans. *Heart Rhythm* 6, 454–460. [PubMed: 19324302]

Nikolaev VO, Moshkov A, Lyon AR, Miragoli M, Novak P, Paur H, Lohse MJ, Korchev YE, Harding SE & Gorelik J. (2010). Beta2-adrenergic receptor redistribution in heart failure changes cAMP compartmentation. *Science* 327, 1653–1657. [PubMed: 20185685]

- Nishisato K, Hashimoto A, Nakata T, Doi T, Yamamoto H, Nagahara D, Shimoshige S, Yuda S, Tsuchihashi K & Shimamoto K. (2010). Impaired cardiac sympathetic innervation and myocardial perfusion are related to lethal arrhythmia: quantification of cardiac tracers in patients with ICDs. *J Nucl Med* 51, 1241–1249. [PubMed: 20679471]
- Norwegian Multicenter Study G. (1981). Timolol-induced reduction in mortality and reinfarction in patients surviving acute myocardial infarction. *N Engl J Med* 304, 801–807. [PubMed: 7010157]
- Packer M, Coats AJ, Fowler MB, Katus HA, Krum H, Mohacs P, Rouleau JL, Tendera M, Castaigne A, Roecker EB, Schultz MK, DeMets DL & Carvedilol Prospective Randomized Cumulative Survival Study G. (2001). Effect of carvedilol on survival in severe chronic heart failure. *N Engl J Med* 344, 1651–1658. [PubMed: 11386263]
- Pare GC, Bauman AL, McHenry M, Michel JJ, Dodge-Kafka KL & Kapiloff MS. (2005). The mA_KAP complex participates in the induction of cardiac myocyte hypertrophy by adrenergic receptor signaling. *J Cell Sci* 118, 5637–5646. [PubMed: 16306226]
- Pauza DH, Pauziene N, Pakeltyte G & Stropus R. (2002a). Comparative quantitative study of the intrinsic cardiac ganglia and neurons in the rat, guinea pig, dog and human as revealed by histochemical staining for acetylcholinesterase. *Ann Anat* 184, 125–136. [PubMed: 11936191]
- Pauza DH, Skripka V & Pauziene N. (2002b). Morphology of the intrinsic cardiac nervous system in the dog: a whole-mount study employing histochemical staining with acetylcholinesterase. *Cells Tissues Organs* 172, 297–320. [PubMed: 12566631]
- Pereira L, Bare DJ, Galice S, Shannon TR & Bers DM. (2017). beta-Adrenergic induced SR Ca(2+) leak is mediated by an Epac-NOS pathway. *J Mol Cell Cardiol* 108, 8–16. [PubMed: 28476660]
- Pham TV, Sosunov EA, Gainullin RZ, Danilo P Jr. & Rosen MR. (2001). Impact of sex and gonadal steroids on prolongation of ventricular repolarization and arrhythmias induced by I(K)-blocking drugs. *Circulation* 103, 2207–2212. [PubMed: 11331264]
- Piccini JP, Simon DN, Steinberg BA, Thomas L, Allen LA, Fonarow GC, Gersh B, Hylek E, Kowey PR, Reiffel JA, Naccarelli GV, Chan PS, Spertus JA, Peterson ED, Outcomes Registry for Better Informed Treatment of Atrial Fibrillation I & Patients. (2016). Differences in Clinical and Functional Outcomes of Atrial Fibrillation in Women and Men: Two-Year Results From the ORBIT-AF Registry. *JAMA Cardiol* 1, 282–291. [PubMed: 27438106]
- Pothineni NV, Shirazi LF & Mehta JL. (2016). Gender Differences in Autonomic Control of the Cardiovascular System. *Curr Pharm Des* 22, 3829–3834. [PubMed: 27189603]
- Ripplinger CM, Noujaim SF & Linz D. (2016). The nervous heart. *Prog Biophys Mol Biol* 120, 199–209. [PubMed: 26780507]
- Rubart M & Zipes DP. (2005). Mechanisms of sudden cardiac death. *J Clin Invest* 115, 2305–2315. [PubMed: 16138184]
- Salama G & Bett GC. (2014). Sex differences in the mechanisms underlying long QT syndrome. *Am J Physiol Heart Circ Physiol* 307, H640–648. [PubMed: 24973386]
- Salameh A, Frenzel C, Boldt A, Rassler B, Glawe I, Schulte J, Muhlberg K, Zimmer HG, Pfeiffer D & Dhein S. (2006). Subchronic alpha- and beta-adrenergic regulation of cardiac gap junction protein expression. *FASEB J* 20, 365–367. [PubMed: 16352648]
- Schwartz PJ, Priori SG, Spazzolini C, Moss AJ, Vincent GM, Napolitano C, Denjoy I, Guicheney P, Breithardt G, Keating MT, Towbin JA, Beggs AH, Brink P, Wilde AA, Toivonen L, Zareba W, Robinson JL, Timothy KW, Corfield V, Wattanasirichaigoon D, Corbett C, Haverkamp W, Schulze-Bahr E, Lehmann MH, Schwartz K, Coumel P & Bloise R. (2001). Genotype-phenotype correlation in the long-QT syndrome: gene-specific triggers for life-threatening arrhythmias. *Circulation* 103, 89–95. [PubMed: 11136691]
- Simpson P (1983). Norepinephrine-stimulated hypertrophy of cultured rat myocardial cells is an alpha 1 adrenergic response. *J Clin Invest* 72, 732–738. [PubMed: 6135712]
- Soltysinska E, Thiele S, Olesen SP & Osadchii OE. (2011). Chronic sympathetic activation promotes downregulation of beta-adrenoceptor-mediated effects in the guinea pig heart independently of structural remodeling and systolic dysfunction. *Pflugers Arch* 462, 529–543. [PubMed: 21811789]
- Somekawa S, Fukuhara S, Nakaoka Y, Fujita H, Saito Y & Mochizuki N. (2005). Enhanced functional gap junction neofunction by protein kinase A-dependent and Epac-dependent signals downstream of cAMP in cardiac myocytes. *Circ Res* 97, 655–662. [PubMed: 16123333]

- Surdo NC, Berrera M, Koschinski A, Brescia M, Machado MR, Carr C, Wright P, Gorelik J, Morotti S, Grandi E, Bers DM, Pantano S & Zaccolo M. (2017). FRET biosensor uncovers cAMP nano-domains at beta-adrenergic targets that dictate precise tuning of cardiac contractility. *Nat Commun* 8, 15031. [PubMed: 28425435]
- Tan AY, Li H, Wachsmann-Hogiu S, Chen LS, Chen PS & Fishbein MC. (2006). Autonomic innervation and segmental muscular disconnections at the human pulmonary vein-atrial junction: implications for catheter ablation of atrial-pulmonary vein junction. *J Am Coll Cardiol* 48, 132–143. [PubMed: 16814659]
- TenBroek EM, Lampe PD, Solan JL, Reynhout JK & Johnson RG. (2001). Ser364 of connexin43 and the upregulation of gap junction assembly by cAMP. *J Cell Biol* 155, 1307–1318. [PubMed: 11756479]
- Turner NA, Porter KE, Smith WH, White HL, Ball SG & Balmforth AJ. (2003). Chronic beta2-adrenergic receptor stimulation increases proliferation of human cardiac fibroblasts via an autocrine mechanism. *Cardiovasc Res* 57, 784–792. [PubMed: 12618240]
- Vaitkevicius R, Saburkina I, Rysevaite K, Vaitkeviciene I, Pauziene N, Zaliunas R, Schauerte P, Jalife J & Pauza DH. (2009). Nerve supply of the human pulmonary veins: an anatomical study. *Heart Rhythm* 6, 221–228. [PubMed: 19187915]
- Vincent GM, Schwartz PJ, Denjoy I, Swan H, Bithell C, Spazzolini C, Crotti L, Piippo K, Lupoglazoff JM, Villain E, Priori SG, Napolitano C & Zhang L. (2009). High efficacy of beta-blockers in long-QT syndrome type 1: contribution of noncompliance and QT-prolonging drugs to the occurrence of beta-blocker treatment “failures”. *Circulation* 119, 215–221. [PubMed: 19118258]
- Vittorio TJ, Zolty R, Kasper ME, Khandwalla RM, Hirsh DS, Tseng CH, Jorde UP & Ahuja K. (2008). Differential effects of carvedilol and metoprolol succinate on plasma norepinephrine release and peak exercise heart rate in subjects with chronic heart failure. *J Cardiovasc Pharmacol Ther* 13, 51–57. [PubMed: 18287590]
- Wagner S, Dybkova N, Rasenack EC, Jacobshagen C, Fabritz L, Kirchhof P, Maier SK, Zhang T, Hasenfuss G, Brown JH, Bers DM & Maier LS. (2006). Ca²⁺/calmodulin-dependent protein kinase II regulates cardiac Na⁺ channels. *J Clin Invest* 116, 3127–3138. [PubMed: 17124532]
- Wallace AG & Sarnoff SJ. (1964). Effects of Cardiac Sympathetic Nerve Stimulation on Conduction in the Heart. *Circ Res* 14, 86–92. [PubMed: 14104166]
- Weiss JN, Garfinkel A, Karagueuzian HS, Chen PS & Qu Z. (2010). Early afterdepolarizations and cardiac arrhythmias. *Heart Rhythm* 7, 1891–1899. [PubMed: 20868774]
- Wilde AA, Moss AJ, Kaufman ES, Shimizu W, Peterson DR, Benhorin J, Lopes C, Towbin JA, Spazzolini C, Crotti L, Zareba W, Goldenberg I, Kanter JK, Robinson JL, Qi M, Hofman N, Tester DJ, Bezzina CR, Alders M, Aiba T, Kamakura S, Miyamoto Y, Andrews ML, McNitt S, Polonsky B, Schwartz PJ & Ackerman MJ. (2016). Clinical Aspects of Type 3 Long-QT Syndrome: An International Multicenter Study. *Circulation* 134, 872–882. [PubMed: 27566755]
- Xiao RP. (2001). Beta-adrenergic signaling in the heart: dual coupling of the beta2-adrenergic receptor to G(s) and G(i) proteins. *Sci STKE* 2001, re15. [PubMed: 11604549]
- Xie Y, Grandi E, Bers DM & Sato D. (2014). How does beta-adrenergic signalling affect the transitions from ventricular tachycardia to ventricular fibrillation? *Europace* 16, 452–457. [PubMed: 24569900]
- Xie Y, Grandi E, Puglisi JL, Sato D & Bers DM. (2013). beta-adrenergic stimulation activates early afterdepolarizations transiently via kinetic mismatch of PKA targets. *J Mol Cell Cardiol* 58, 153–161. [PubMed: 23481579]
- Yamamoto T, El-Hayek R & Ikemoto N. (2000). Postulated role of interdomain interaction within the ryanodine receptor in Ca(2+) channel regulation. *J Biol Chem* 275, 11618–11625. [PubMed: 10766778]
- Yancy CW, Jessup M, Bozkurt B, Butler J, Casey DE Jr., Colvin MM, Drazner MH, Filippatos GS, Fonarow GC, Givertz MM, Hollenberg SM, Lindenfeld J, Masoudi FA, McBride PE, Peterson PN, Stevenson LW & Westlake C. (2017). 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. *J Card Fail* 23, 628–651. [PubMed: 28461259]

- Yancy CW, Jessup M, Bozkurt B, Butler J, Casey DE Jr., Drazner MH, Fonarow GC, Geraci SA, Horwich T, Januzzi JL, Johnson MR, Kasper EK, Levy WC, Masoudi FA, McBride PE, McMurray JJ, Mitchell JE, Peterson PN, Riegel B, Sam F, Stevenson LW, Tang WH, Tsai EJ, Wilkoff BL, American College of Cardiology F & American Heart Association Task Force on Practice G. (2013). 2013 ACCF/AHA guideline for the management of heart failure: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. *J Am Coll Cardiol* 62, e147–239. [PubMed: 23747642]
- Zaccolo M (2009). cAMP signal transduction in the heart: understanding spatial control for the development of novel therapeutic strategies. *Br J Pharmacol* 158, 50–60. [PubMed: 19371331]
- Zaccolo M & Pozzan T. (2002). Discrete microdomains with high concentration of cAMP in stimulated rat neonatal cardiac myocytes. *Science* 295, 1711–1715. [PubMed: 11872839]
- Zaglia T & Mongillo M. (2017). Cardiac sympathetic innervation, from a different point of (re)view. *J Physiol* 595, 3919–3930. [PubMed: 28240352]
- Zhou J, Shin HG, Yi J, Shen W, Williams CP & Murray KT. (2002). Phosphorylation and putative ER retention signals are required for protein kinase A-mediated potentiation of cardiac sodium current. *Circ Res* 91, 540–546. [PubMed: 12242273]
- Zhou J, Yi J, Hu N, George AL Jr. & Murray KT. (2000). Activation of protein kinase A modulates trafficking of the human cardiac sodium channel in *Xenopus* oocytes. *Circ Res* 87, 33–38. [PubMed: 10884369]

Highlights

- β -blockers are effective antiarrhythmics in many conditions, including heart failure, myocardial infarction, and atrial fibrillation
- β -adrenergic action on cardiac function is complex, and involves structural and functional changes occurring over multiple time- and spatial-scales.
- Exploiting drugs' receptor-specific drug action, targeting compartmentalized signaling, and understanding sex differences in drug responses are key aspects that future studies should address to improve personalization of β -blockade therapy to varying arrhythmia types and patient groups.

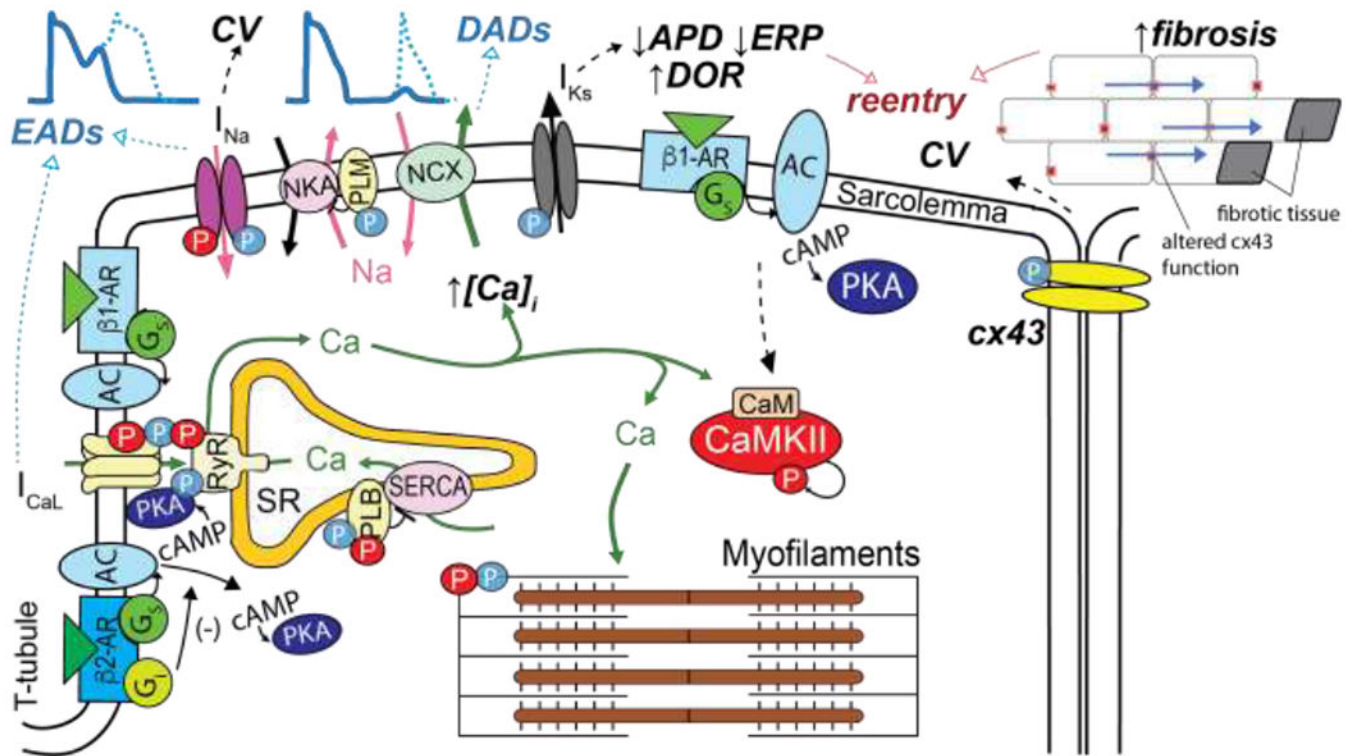


Figure 1: Molecular and cellular mechanisms of β -adrenergic action.

Schematic of the main cellular processes linking β -adrenergic activation (and β -adrenergic receptor specific signaling) to increased propensity for arrhythmia. Protein kinase A (PKA) phosphorylates multiple targets (blue symbol "P") that directly affect membrane electrophysiology and Ca signals. These include the L-type Ca channel (carrying I_{CaL}), ryanodine receptor (RyR), phospholamban (PLB) that regulates the sarcoplasmic reticulum Ca pump (SERCA), phospholemman (PLM) that regulates the Na/KATPase (NKA), the sarcolemmal Na and K channels (carrying I_{Na} and I_{Ks}), myofilament proteins, and connexin 43 (cx43). These downstream effects of β -adrenergic action facilitate the development of ectopic activity (EADs and DADs) and functional reentry (shortened action potential duration, APD and effective refractory period, ERP, increased dispersion of repolarization, DOR, and altered conduction, CV). β -adrenergic activation is also involved in structural remodeling (e.g., fibrosis) that facilitates the formation of a structural reentrant substrate. CaMKII is a central player in cardiac disease and adrenergically-mediated arrhythmia; its activity is enhanced by β -adrenergic activation via both increases in Ca and cAMP, and leads to increased phosphorylation of many of the same PKA targets (red symbol "P").

**Table 1 –
Mechanisms of action and therapeutic uses of β -blockers.**

Currently used β -blockers, their mechanisms of action (including β -AR receptor sensitivity), other extra-cardiac actions, and therapeutic uses are listed. Specific indications for arrhythmia are in red, and indication for HF, MI and AF are in blue. Information compiled from (Brunton et al., 2018).

Agent	Mechanism of Action	Therapeutic use
Non-selective β-adrenergic antagonists (first generation):		
Propranolol	Equal affinity for β_1 and β_2 . Membrane stabilizing effect.	Used for: hypertension, angina, supraventricular arrhythmia, ventricular arrhythmia, MI.
Nadolol	Equal affinity for β_1 and β_2 . No sympathomimetic or membrane stabilizing activity.	Used for: Hypertension, angina, LQTS.
Timolol	Equal affinity for β_1 and β_2 . No sympathomimetic or membrane stabilizing activity.	Hypertension, congestive HF, acute MI.
β_1-selective adrenergic antagonists (second generation):		
Metoprolol	No sympathomimetic or membrane stabilizing activity.	Used for: essential hypertension, angina, tachycardia, HF, vasovagal syncope, secondary prevention after MI
Atenolol	No sympathomimetic or membrane stabilizing activity.	Used for: hypertension, coronary heart disease, arrhythmias, angina, reduces risk of complications after MI
Esmolol	Little sympathomimetic activity, no membrane-stabilizing activity.	Used when short duration is desired or in critically ill patients where rapid withdrawal may be necessary.
Acebutolol	Some sympathomimetic and membrane stabilizing activity.	Used for hypertension, atrial and ventricular arrhythmias, acute MI in high-risk patients
Bisoprolol	No sympathomimetic or membrane stabilizing activity. Higher degree of β_1 selectivity than metoprolol or atenolol.	Used for: HF, hypertension, MI, arrhythmias
β-adrenergic antagonists with additional cardiovascular effects (third generation - also possess vasodilatory actions)		
Labetalol	Competitive antagonist to α_1 and β receptors (β_1 and β_2). Partial agonist activity at β_2 and also inhibits neuronal uptake of NE (cocaine-like).	Used for chronic hypertension or hypertensive emergencies
Carvedilol	Blocks α_1 , β_1 , and β_2 similar to labetalol, but also has anti-oxidant and anti-inflammatory properties. Has membrane-stabilizing action, but no sympathomimetic activity.	Produces vasodilation and anti-inflammatory effects may help treatment of HF. Approved for use in hypertension, congestive HF, and LV dysfunction after MI
Celiprolol	β_1 antagonist. β_2 partial agonist. Also α_2 antagonist and promotes NO production.	Reduces HR and blood pressure. Used to treat hypertension and angina
Nebivolol	β_1 antagonist with endothelial NO-mediated vasodilatory action.	Also has antioxidant action and neutral or favorable effects on carbohydrate and lipid metabolism. Approved for the treatment of hypertension